Sesquiterpenoid Constituents of the Liverwort, Ptuchanthus striatus (LEHM. et LINDENB.) NEES

Reiji TAKEDA,* Hideo NAOKI, Takashi Iwashita, Kosei Mizukawa, Yoshio Hirose, Toshimasa Isida,† and Masatoshi Inoue†

Suntory Institute for Bioorganic Research, Wakayama-dai, Shimamoto-cho, Mishima-gun, Osaka 618

†Osaka College of Pharmacy, 2-10-65 Kawai, Matsubara, Osaka 580

(Received October 25, 1982)

Four sesquiterpenoids, striatene, striatol, β -monocyclonerolidol and ptychanolide, were isolated from the liverwort *Ptychanthus striatus* (Lehm. et Lindenb.)Nees. Their structures have been established by spectroscopic analysis and chemical transformation. Three of them, striatene, striatol and β -monocyclonerolidol, are interesting in terms of the evolution of the liverwort from the algae.

Previously, a number of compounds have been isolated from the liverwort belonging Jungermanniales.1) In the course of our investigation on the terpene constituents of the liverwort, we examined the constituents of Ptychanthus striatus (LEHM. et LINDENB.) NEES and isolated four new sesquiterpenes, striatene, striatol, β -monocyclonerolidol and ptychanolide together with two known sesquiterpenes, deoxopinguisone²⁾ and pinguisanene.³⁾ Three of them, striatene, striatol and β -monocyclonerolidol are interesting in terms of the evolution of the liverwort and the algae. Asakawa et al.3) have reported the identification of mono- and sesquiterpene hydrocarbons, e.g. α-pinene, β -pinene, camphene, calamenene, and α -copaene, from the same liverwort. This paper deals in detail with the structural determination of striatene, striatol, β monocyclonerolidol4) and ptychanolide.5)

Striatene (1), striatol (2), and ptychanolide (4) were isolated from the acetone extract of the dry material collected in Tokushima Prefecture in August 1979 by column chromatography on SiO_2 using hexane (F-1) and $CHCl_3$ (F-2 and F-3), while striatene (1), β -monocyclonerolidol (3), and ptychanolide (4) were isolated from the dry material collected in Nara Prefecture in November 1979 by similar procedure.

Striatene (1), $[\alpha]_D + 72.7^\circ$, was obtained as a colorless oil by preparative GC of the hydrocarbon fraction. The molecular weight by HR-MS together with the ¹H and ¹³C NMR data indicated the molecular formula to be $C_{15}H_{24}$. Its IR (1600 cm⁻¹) and UV (λ_{max} 238 nm, ε

24400) spectra showed the presence of conjugated diene system. The IR (1640 and 990 cm⁻¹) and ¹H NMR (δ 5.25, 5.28, and 6.91) spectra also showed characteristic bands and signals due to vinyl groups. Moreover, the ¹H NMR spectrum showed signals due to a tertiary methyl at δ 0.96, a secondary methyl at δ 0.92, two olefinic methyls at δ 1.69 and 1.88, two trisubstituted double bonds at δ 5.34 and 5.56, and an allylic methylene at δ 2.18 and 2.49.

Striatol (2), $[\alpha]_D$ +49.5°, was also obtained as a colorless oil by rechromatography on AgNO₃-SiO₂ of F-2. The molecular formula C₁₅H₂₆O of 2 was determined by HR-MS together with the ¹H and ¹³C NMR data. The IR (3400, 1640, 990, and 880 cm⁻¹) spectrum showed bands due to hydroxyl, trisubstituted double bond and vinyl groups. The olefinic functions were further supported by ${}^{1}H$ [δ 5.41 (3-H), 5.09, 5.26 (11- $H_{A,B}$), and 5.92 (10-H)], and ¹³C NMR (δ 124.0, 145.1, 139.3, and 111.4) spectra. Moreover, ¹H NMR spectrum showed signals due to a secondary methyl at δ 0.87, two tertiary methyls at δ 0.88 and 1.29, and an olefinic methyl at δ 1.59. Treatment of 2 with POCl₃ in dry pyridine yielded two trienes, one of which was identified as striatene (1) by GLC, IR, and MS. The above spectral data together with this result indicated that striatene and striatol have the same monocyclic carbon skeleton. The presence of the partial structure 2a was indicated by the results of double irradiation experiments in the ¹H NMR spectrum of 2 using Eu(fod)₃. Irradiation of the signal of 6-CH₃ converted a triplet of quartets of 6-H into a triplet (J=6.1 Hz). The fact that 6-H is coupled to 1α - and 1β -H's with J=6.1 Hz indicates it is equatorial. Also, irradiation of 1-HA collapsed the multiplet of 2-HA to a sharp peak and a triplet of quartets of 6-H to a doublet of quartets. Further, irradiation of the signal due to 2-H_A collapsed a broad singlet due to 3-H and a multiplet due to $1-H_{\blacktriangle}$ to sharp peaks.

Striatene (1) was oxidized with *m*-chloroperbenzoic acid (MCPBA) in CH_2Cl_2 at 0 °C to give two monoepoxides **5** and **6**, the UV spectra of which indicated absorption maximum at 238 nm (ε 19500) due to a conjugated diene. The ¹H NMR spectra of epoxides **5** and **6** indicated signals of four olefinic protons (**5**: δ 5.08, 5.21, 5.35, and 6.73; **6**: 5.06, 5.20, 5.67, and 6.82) and a methine proton (**5**: δ 3.01; **6**: 2.77) attached to an epoxy ring. These results indicated that the double

bonds at C₃/C₄ was epoxidized. The stereochemistry of oxirane rings in 5 and 6 were determined by ¹H NMR using shift reagent, i.e., two protons of α -epoxide 6 at C-7 underwent a much larger shift than those of β epoxide 5, thus showing that the oxirane ring and side chain in 6 have a cis relation, while those of in 5 have a trans relation. The presence of the C_6 side chain was suggested by the mass spectra of 1 and 5, and ¹H NMR of 5. Namely, the mass spectra contained intense ions at m/z 123 (M-81) for 1, m/z 139 for 5 and m/z 81 for both compounds originating by cleavage of the C-5/C-7 bond. Furthermore, in the ¹H NMR spectrum of 5, irradiation at δ 5.21 (8-H) collapsed the allylic methylene at δ 2.15 (7-H_A) and 2.41 (7-H_B) to AB type doublets and the olefinic methyl protons (9-CH₃) at $\delta = 1.86$ to a sharp peak. These results indicated that the C₆ side chain in 1 is 3-methyl-1,3-pentadiene. In the ¹H NMR of 2 with the aid of shift reagents [Eu(fod)₃], the proton at C-6 together with the CH₃ group at C-4 were observed to have larger shifts than CH₃ groups at C-5 and C-6. In the NOE experiments of benzoate 9 (described below), a 2.7% NOE was observed on the 6-CH₃ group (axial) upon irradiation of 5-CH₃.6) These results indicated that 6-H and the hydroxyl-containing side chain have a cis relation. The geometry of 8-ene in 1 was determined by NOE experiment. An 11% NOE was observed on 8-H upon irradiation of 9-CH₃, i.e., the 8-ene is Z. These results lead to structures 1 and 2 for striatene and striatol, respectively.

The absolute configurations of 1 and 2 were determined in the following way. Hydrogenation of monoepoxide 5 with Pd/C gave tetrahydro compound 7, the IR spectrum of which showed the absence of double bond absorption. The ¹H NMR spectrum indicated

signals of two secondary and one primary methyl groups at δ 0.74, 0.84, and 0.88. Compound 7 was treated with diethylamine and n-BuLi7) in dry ether to give 8 which was then reacted with p-bromobenzoyl chloride in dry pyridine to yield the corresponding monobenzoate 9. The IR spectrum exhibited bands at 1718, 1640, and 890 cm⁻¹ due to an ester group and an exocyclic methylene group. The ¹H NMR spectrum indicated the presence of a methine proton adjacent to an oxygen (δ 5.57), exocyclic methylene protons (δ 4.76 and 5.08) and protons on an aromatic ring (δ 7.59 and 7.97). The fact that 3-H is coupled to 2-H_{A.B} with J=5.3 and 10.9 Hz indicates that it is axial. The signal of 6-H at δ 1.67 appeared as a triple quartet, (J=4.8 and 6.6 Hz) which collapsed to triplet (J=4.8 m)Hz) on irradiation of 6-CH₃. This fact indicates that 6-H is equatorial. In the NOE experiments of 9, an 11% NOE is observed on 12-H_A upon irradiation of 5-CH₃, i.e., 5-CH₃ is equatorial. From these results the conformation of this compound was determined to be as shown by 9a. Harada et al. reported that the absolute configuration of cyclic allylic alcohols can be determined nonempirically by the CD exciton chirality method.8) Application of this method to benzoate 9, $\Delta \varepsilon_{241} + 3.1$, shows that the exocyclic double bond and the 3-OCOPh group constitute a positive chirality as Thus, the absolute configuration of shown in 9b. striatene is as shown in structure 1.

The configuration of the t-OH group in 2 was determined by taking (R)-(-)-linalool (10) as the reference sample. Namely, it was found that the p-bromobenzoates of striatol and (R)-(-)-linalool both show negative Cotton effects at 252 nm (in MeOH), 2b $\Delta \varepsilon_{252}$ -0.4 and 10a $\Delta \varepsilon_{252}$ -0.5. This establishes the C-9 configuration in 2 to be R. Recently, Gonnella et al.9) have shown that the benzoate method described above for cyclic compounds is extensible to acyclic secondary allylic alcohols, namely, that the benzoate of acyclic allylic moiety 11 exhibits a positive CD. The present results including that of linalool show that the method is applicable to t-OH system 12 as well (because the methyl group is smaller than other alkyl substituents).

 β -Monocyclonerolidol (3), $[\alpha]_D + 3.2^\circ$, was obtained as a colorless oil by rechromatography on AgNO₃-SiO₂

Chart 3.

The molecular formula C₁₅H₂₆O of 3 was determined by HR-MS together with the ¹H and ¹³C NMR data. The IR spectrum showed the presence of a hydroxyl group at 3400 cm⁻¹, a vinyl group at 1645 and 990 cm⁻¹, and an exocyclic methylene group at 890 cm⁻¹. The ¹H NMR spectrum indicated signals of gem-dimethyl groups at δ 0.84 and 0.92, a tertiary methyl group on a carbon bearing hydroxyl at δ 1.27, an exocyclic methylene group at δ 4.53 and 4.75, and vinyl groups at δ 5.02, 5.18, and 5.91. The mass spectrum indicated intense ion peaks at m/z 123 (M-18-81) and m/z 81 as observed in 1 and 2. The fact indicates that 3 also has a C6 side chain. From these results together with biogenetic consideration, we assumed that 3 had a monocyclonerolidol skeleton as shown by structure 3. This assumption was confirmed by partial synthesis of 3 from α -ionone.

Hydrogenation of commercial (+)- α -ionone 13 with Pd/C in 1.5% KOH-EtOH gave the hydrogenated mixture, from which dihydroionone 14 was separated by column chromatography on SiO₂. Dihydroionone 14 exhibited the molecular ion peak at m/z 194 in its mass spectrum. The IR spectrum exhibited bands at 1710 and 810 cm⁻¹ due to a carbonyl group and a trisubstituted double bond. The ¹H NMR spectrum indicated the presence of an olefinic methyl group (δ 1.66) and a trisubstituted double bond (δ 5.31). The reaction of 14 with ethylene glycol and TsOH yielded acetal 15, the ¹H NMR spectrum of which exhibited a signal due to the acetal group at δ 3.90. The IR spectrum showed the absence of a carbonyl group. Next, irradiation of acetal 15 in ether containing 10% phenol with a 450 W-Hg lamp¹⁰⁾ afforded a mixture of 16 and starting material 15. Then, this mixture was hydrolyzed with TsOH in THF containing H₂O to give a ketone mixture, 16a and 14, from which the exocyclic isomer 16a was separated by column chromatography on AgNO₃-SiO₂. The IR spectrum showed exocyclic methylene absorption bands at 1640 and 885 cm⁻¹, and a carbonyl absorption band at 1710 cm⁻¹. The ¹H NMR spectrum also suggested the presence of an exocyclic methylene group at δ 4.49 and 4.74. The exocyclic isomer 16a was reacted with vinylmagnesium bromide (prepared from vinyl bromide and magnesium in THF) in dry THF to give the racemic alcohol 17. The NMR, IR, and mass spectra of 17 were identical with those of β -

a) H₂, pd/C, 1.5% KOH-EtOH; b) Ethylene glycol, TsOH c) 10% Penol/ether, $h\nu(450 \text{ W})$; d) TsOH, THF THF; e) CH₂=CHMgBr, THF.

monocyclonerolidol (3).

On the basis of distribution of terpenoids, Asakawa and co-workers¹¹⁾ have noted that liverwort are closely related to algae. The fact that the skeletal structures of 1-3 are identical with α - and β -snyderol,¹²⁾ microcionin,¹³⁾ etc.¹⁴⁾ which have been found in marine algae and in marine animals feeding on algae, supports the notion that liverworts have evolved from algae.

Ptychanolide (4), mp 143—144 °C, $[\alpha]_D$ +23.2° was obtained as a colorless needles by recrystallization of F-2 and 2'. The molecular formula C₁₅H₂₂O₃ of 4 was determined by the appearance of a molecular ion peak at 250.1565 in the HR-MS. The IR spectrum showed the presence of a y-lactone absorption band at 1780 cm⁻¹ and the absence of hydroxyl absorptions. The ¹H NMR spectrum indicated signals due to two secondary CH₃ groups at δ 0.84 and 0.98, two tertiary CH₃ groups at δ 0.79 and 0.93, an isolated methylene group at δ 1.39 and 1.99, and two methine protons adjacent to oxygen at δ 3.55 and 5.52. The ¹³C NMR spectrum also suggested the presence of a γ -lactone group at δ 76.1 and 179.8. Hence the remaining oxygen atom was attributed to an ether linkage. Treatment of the lactone 4 with 10% H₂SO₄ in acetone afforded the diol lactone 18. The IR spectrum showed bands at 3500 and 1770 cm⁻¹ due to hydroxyl and γ -The ¹H NMR spectrum indicated lactone groups. signals of two methine protons adjacent to oxygen at δ 4.13 and 5.62 which were coupled with each other. The formation of an a-glycol from an ether indicates that the diol is derived from an epoxide. The lactone 4 was reduced with LAH in dry ether to triol 19, the ¹H NMR spectrum of which showed signals of five protons adjacent to oxygen at δ 3.64, 3.72, 3.76, 3.84, and 3.92. In decoupling experiments, irradiation at δ 3.92 collapsed the two double doublets of the -CH₂OH group at δ 3.72 and 3.76 to AB type doublets. This result and assignments of J values suggested that a hydroxymethyl group at C-11 and 1,2-diol system at C-9 and C-10 are attached to a quaternary carbon. Hence, the γ -lactone must form a spiro linkage at C-6 and the epoxy ring is attached to the γ -lactone ring.

Oxidation of triol 19 with NaIO₄ gave aldehyde 20, M^+ m/z 224 ($C_{14}H_{24}O_2$), the IR spectrum of which showed bands at 3560, 3450, and $1715~\mathrm{cm^{-1}}$ due to hydroxyl and formyl group. The ¹H NMR spectrum indicated signals of two protons adjacent to an oxygen as primary alcohol at δ 3.55 and 3.94, and proton of formyl group at δ 9.69 instead of three protons adjacent to oxygen in 19. Aldehyde 20 was further oxidized with MCPBA in CH2Cl2, followed by hydrolysis with Al_2O_3 to yield diol 21 (M+, m/z 212; $C_{13}H_{24}O_2$). Its IR spectrum showed the presence of a hydroxyl absorption band at 3325 cm⁻¹ and the absence of carbonyl absorption. The diol 21 was oxidized further with NaIO₄ to the corresponding ketone 22 which exhibited a molecular ion peak at 180 (C₁₂H₂₀O) in the mass spectrum. The IR spectrum indicated the presence of a five-membered ring ketone (1740 cm⁻¹). These results establish that ptychanolide 4 has a spiro lactone moiety at C-6. Reduction of ketone 22 with LAH in dry ether afforded alcohols 23 and 24, the ¹H NMR

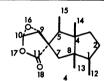
spectra of which showed signals of protons on carbons bearing hydroxyl groups (23: δ 3.77; 24: 4.20), respectively. In the decoupling experiments of 23, irradiation of 6-H as a double triplet collapsed a pair of double doublets of 7-CH₂ at δ 1.49 and 1.79 to a pair of doublets and a double quartet of 5-H at δ 1.74 to a quartet. This fact indicated that the OH function was flanked by a methylene group and secondary CH₃ group. Namely, ptychanolide has a partial structure 4a. Ketone 22 was treated with 1,2-ethanedithiol to give the corresponding thioacetal which was desulfurized with Raney Ni in EtOH to yield the volatile hydrocarbon 25. Its mass spectrum showed a molecular ion peak at 166 (C₁₂H₂₂). However, the ¹H and ¹³C NMR spectra showed only the signals of two methyl groups and five protons, and those of six carbons, respectively. Therefore, the hydrocarbon 25 has the symmetric bicyclo[3.3.0]octane skeleton. The above results indicate that ptychanolide is represented by formula 4 (planar).

Chart 5.

The stereostructure of ptychanolide (4) was determined by X-ray analysis. The colorless transparent crystals of 4, which were recrystallized from hexane–EtOAc, belong to orthorhombic with space group $p2_12_12_1$. The crystal data are as follows: $C_{15}H_{22}O_3$, $M_r=250.16$, a=6.514(1), b=19.376(4), c=10.543(3) Å, v=1330.7(5) ų, Z=4, $D_m=1.233$ (1), $D_x=1.249$ g cm⁻³. X-Ray diffraction intensities were measured by Rigaku automatic diffractometer with graphite-monochromated Cu $K\alpha$ radiation using a θ -2 θ scan mode and a scan rate of 4° /min. Stationary background counts (5 s each) were taken at both limits of each scan (scan width in 2θ : 1.2+0.15 tan θ). A total of 1344 unique reflections were measured to the limit $2\theta=130^\circ$. Lorentz and polarization corrections were applied, but no absorption

Table 1. Atomic coordinates (\times 10⁴) of nonhydrogen atoms of 4

Atom	x	у	z
C1	4160 (11)	977 (3)	5420 (6)
C2	3997 (11)	237 (3)	5870 (8)
C3	5405 (11)	162 (4)	7049 (8)
C4	6291 (8)	886 (3)	7297 (6)
C 5	4842 (9)	1322 (3)	8111 (6)
C 6	5179 (8)	2073 (3)	7648 (6)
C 7	6138 (9)	2006 (3)	6283 (6)
C8	6217 (8)	1238 (3)	5977 (5)
C 9	6445 (9)	2549 (3)	8451 (6)
C10	5357 (11)	3181 (4)	8584 (6)
C11	3183 (9)	2479 (3)	7560 (6)
C12	3899 (12)	1056 (4)	3983 (8)
C13	8035 (10)	1071 (4)	5107 (7)
C14	8450 (10)	842 (4)	7867 (7)
C15	4977 (12)	1232 (4)	9551 (7)
O16	5529 (7)	2767 (3)	9630 (4)
O17	3428 (7)	3132 (2)	7980 (5)
O18	1566 (6)	2293 (2)	7122 (5)



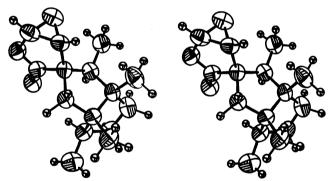


Fig. 1. Perspective drawing of ptychanolide 4.

correction was made because of the small size of the used crystal (dimension: $0.3 \times 0.4 \times 0.2 \text{ mm}^3$). The X-ray intensities of four standard reflections monitored at 100 reflection intervals showed no evidence of structural deterioration during the data collection.

The structure was solved by the direct methods with program MULTAN78. An E-map, calculated using 200 reflections (|E|>1.40) with the phase set of the highest combined figure of merit (=2.86), revealed the positions of all nonhydrogen atoms. These coordinates were refined by a full-matrix least-squares method with isotropic temperature factors and then by a block-diagonal least-squares method with anisotropic ones. All hydrogen atoms could be located from a difference Fourier map and these were included in a further refinement with isotropic temperature factors. The final R value was 0.078. The final positional parameters with their estimated standard deviations of nonhydrogen atoms are listed in Table 1.

The atomic scattering factors for all atoms were taken from "International Tables for X-Ray Crystallography."16) All numerical calculations were carried out on an ACOS-700 computer at the Computation Center of Osaka University using UNICS program. 17) The stereostructure of 4 is shown in Fig. 1. The relative stereostructure of ptychanolide (4) was thus clarified. Furthermore, the keto compound 22 showed a positive Cotton effect at 295 nm in the CD spectrum (Fig. 2). The positive sign of the Cotton effect can be understood by application of the octant rule¹⁸⁾ to the cyclopentanone system, as visualized in octant projection formula 22 This leads to the absolute configuration and **22a**. shown for 22 which is corroborated by the established configuration of congeners 26^{2,19} and 27.3 Thus, the absolute structure of ptychanolide is shown by formula

The biogenesis of the pinguisone type sesquiterpenoids represented by deoxopinguisone 26 and pinguisanene 27 is as yet not clear. However, the isolation of ptychanolide 4 from the same liverwort suggests that they are derived from an intermediate such as 28.

Experimental

All melting points are uncorrected. IR spectra were

measured with a Hitachi EP1-G2 spectrometer, ¹³C NMR spectra with a JEOL FX-100 (25.0 MHz) spectrometer, ¹H NMR spectra with a JEOL FX-100 (100 MHz), a Nicolet NT-360 (360 MHz) or a Hitachi R-20B (60 MHz) spectrometer in deuteriochloroform solution containing tetramethylsilane as an internal standard, low resolution mass spectra with a Hitachi RMU-6 mass spectrometer and high resolution mass spectra with a JEOL 01SG-2, with direct inlet system operating at 70 eV. CD spectra were measured with a JASCO J-20 spectrometer. An analytical GC was performed with a Hitachi 163-type apparatus equipped with a Thermon 600T glass capillary column, and preparative GC with Varians model 90 fitted OV-17 on Chromosorb-W, operated at 150°. Kieselgel 60 (E. Merck, Darmstadt) and Silica CC-7 special (Mallinckrodt) were used for column chromatography. Thin-layer chromatography (TLC) was carried out on Kieselgel GF₂₅₄ (E. Merck, Darmstadt) in 0.25 mm thickness.

Extraction and Isolation. The dry material (100 g) of Ptychanthus striatus (Lehm. et Lindenb.) Nees collected in Wakayama Prefecture around Kosa-cho in March 1980 was extracted with acetone at room temperature. Acetone was evaporated under vacuum and the concentrate was extracted with CHCl₃. The CHCl₃ extract (6.6 g) was subjected to column chromatography over SiO₂. The fraction eluted with hexane gave sesquiterpene hydrocarbons (F-1) containing striatene (1), deoxopinguisone (26), and pinguisanene (27). The fraction eluted with CHCl₃ yielded crude ptychanolide (4) (F-2) and crude striatol (2) (F-3). Also, F-1', F-2', and F-3' were isolated from the acetone extract of the dry material (100 g) collected in Nara Prefecture around Ikadaba in November 1979 by a similar procedure.

Striatene (1) was isolated from hydrocarbon fractions, F-1 and F-1', by preparative GC, as a colorless oil (1, 200 mg from F-1 and F-1', respectively), $[a]_D^{22} + 72.7^\circ$ (c 1.19, CHCl₃), IR (film) 1640, 1600, 990, 810 cm⁻¹; UV (cyclohexane) 232 (sh), 238 (ε 24400) and 245 (sh) nm; ¹H NMR δ 0.92 (3H, d, $J=6.5 \text{ Hz}, 6-\text{CH}_3$), 0.96 (3H, s, 5-CH₃), 1.69 (3H, br.s, 4- CH_3), 1.88 (3H, br.s, 9- CH_3), 2.18 (1H, dd, J=6.0, 16.0 Hz, $7-H_A$), 2.49 (1H, dd, J=8.0, 16.0 Hz, $7-H_B$), 5.25 (1H, dd, J=1.6, 10.5 Hz, 11-H_A), 5.28 (1H, dd, J=1.6, 17.3 Hz, 11-H_B), 5.34 (1H, m, 8-H), 5.56 (1H, t-like, 3-H), and 6.91 (1H, dd, J=10.5, 17.3 Hz, 10-H); ¹³C NMR δ 16.0 (q), 19.2 (q), 20.0 (q), 20.7 (q), 25.3 (t), 27.1 (t), 34.2 (t), 34.2 (d), 40.7 (s), 113.2 (t), 124.4 (d), 128.1 (d), 134.0 (d), 133.2 (s), and 139.1 (s); MS m/z (%) 204 (7, M+), 123 [100, (M-81)+] and 81 (25). Found: m/z 204.1866. Calcd for $C_{15}H_{24}$: M, 204.1856.

Epoxidation of Striatene (1). The solution of m-chloroperbenzoic acid (390 mg) in CH_2Cl_2 (5 ml) was added to an ice-cooled solution of 1 (300 mg) in CH_2Cl_2 (5 ml) and the mixture was stirred for 40 min at 0 °C. The reaction mixture was treated in the usual way to give residue (190 mg), which was chromatographed on 10% AgNO₃-SiO₂ (Mallinckrodt, CC-7). Elution with CH_2Cl_2 -EtOAc (20:1) gave β -epoxide 5 (55 mg) as a colorless oil. Successive elution with the same solvent gave α -epoxide 6 (10 mg) as a colorless oil.

β-Epoxide 5: IR (film) 1640, 1600, 980, 900 cm⁻¹; UV (cyclohexane) 237 nm (ε 19500); ¹H NMR δ 0.75 (3H, d, J=6.5 Hz, 6-CH₃), 0.92 (3H, s, 5-CH₃), 1.26 (3H, s, 4-CH₃), 1.86 (3H, br.s, 9-CH₃), 2.15 (1H, dd, J=5.5, 17.0 Hz, 7-H_A), 2.41 (1H, dd, J=7.5, 17.0 Hz, 7-H_B), 3.01 (1H, br.s, 3-H), 5.08 (1H, dd, J=1.5, 10.5 Hz, 11-H_A), 5.21 (1H, t-like, 8-H), 5.35 (1H, dd, J=1.5, 17.5 Hz, 11-H_B), and 6.73 (1H, dd, J=10.5, 17.5 Hz, 10-H); MS m/z (%) 220 (3, M⁺, C₁₈H₂₄O), 205 [8, (M-CH₃)⁺], 139 [98, (M-81)⁺], 109 (92), 95 (100), 81 (99), 43 (98).

α-Epoxide 6: IR (film) 1645, 1605, 980, 900 cm⁻¹; UV

(cyclohexane) 238 nm (ε 18000); ¹H NMR δ 0.72 (3H, d, J=6.5 Hz, 6-CH₃), 0.91 (3H, s, 5-CH₃), 1.27 (3H, s, 4-CH₃), 1.86 (3H, br.s, 9-CH₃), 2.36 (2H, br.s, 7-H_{A.B}), 2.77 (1H, t, J=1.5 Hz, 3-H), 5.06 (1H, dd, J=1.5, 10.5 Hz, 11-H_A), 5.20 (1H, dd, J=1.5, 17.5 Hz, 11-H_B), 5.67 (1H, br.t, J=4.0 Hz, 8-H), and 6.82 (1H, dd, J=10.5, 17.5 Hz, 10-H); MS m/z (%) 220 (trace, M+, C₁₅H₂₄O), 139 [17, (M-81)+], 95 (30), 81 (20), 43 (100).

Hydrogenation of β-Epoxide 5. β-Epoxide 5 (10 mg) and 10% palladium charcoal (10 mg) in EtOH (1 ml) were stirred under hydrogen atmosphere at room temperature for 5 h. The catalyst was removed by filtration and the solvent was evaporated to give a residue (11 mg), which was chromatographed on SiO₂. Elution with CHCl₃ yielded tetrahydro epoxide 7 (7 mg), colorless oil, ¹H NMR δ 0.74 (3H, d, J=6.5 Hz, s-CH₃), 0.84 (3H, d, J=7.5 Hz, s-CH₃), 0.86 (3H, s, 5-CH₃), 0.88 (3H, t, J=6.6 Hz, 10-CH₃), 1.24 (3H, s, 4-CH₃), and 2.96 (1H, br.s, 3-H); MS m/z (%) 224 (12, M⁺, C₁₅H₂₈O), 209 [31, (M-CH₃)⁺], 103 (100), 85 (98), 43 (76).

Conversion of 7 into Benzoate 9. Commercial butyllithium (0.3 ml, 15% solution in hexane) was added to a solution of diethylamine (36.5 mg) in dry ether (1 ml). Tetrahydro epoxide 7 (18 mg) in dry ether (1 ml) was then added and the mixture was refluxed for 3 h. The reaction mixture was poured into NaCl aqueous solution and extracted with ether. The ether layer was treated in the usual way to give residue (14 mg), which was treated with p-bromobenzoyl chloride (20 mg) in dry pyridine (2 ml) at room temperature for a day. The reaction mixture was treated in the usual way to give a residue, which was chromatographed on SiO₂. Elution with hexane-EtOAc (20:1) gave benzoate 9 (8 mg), colorless viscous oil, IR (film) 1718, 1640, 890 cm⁻¹; UV (MeOH) 205.5, 244 nm (ε 16900, 17500); CD, $\Delta \varepsilon_{241}$ +3.06 (ε 0.145, MeOH); ¹H NMR δ 0.82 (3H, t, J=6.6 Hz, 10-CH₃), 0.83 (3H, d, J=6.6 Hz, 9-CH₃), 0.90 (3H, d, J=6.7 Hz, 6-CH₃), 1.01 (3H, s, 5-CH₃), 1.67 (1H, tq, J=4.8, 6.6 Hz, 6-H), 4.76 (1H, br.s, $12-H_A$), 5.08 (1H, br.s, $12-H_B$), 5.57 (1H, dd, J=5.3, 10.9 Hz, 3-H), 7.59 (2H, aromatic protons) and 7.98 (2H, aromatic protons); MS m/z (%) 408, 406 (trace, M+, C₂₂H₃₁O₂Br), 323, 321 [45, 47, (M-85)+], 185, 183 [92, 91, (p-BrC₆H₄CO)+], 122 (100), 93 (69).

Benzoylation of (R)-(-)-Linalool (10). The mixture of (R)-(-)-linalool (10) (20 mg) isolated from Ho leaf oil and p-bromobenzoyl chloride (20 mg) in dry pyridine (1 ml) was kept at room temperature for 2 d. The reaction mixture was treated in the usual way to yield a residue, which was chromatographed on SiO₂. Elution with CHCl₃ gave a benzoate 10a (15 mg), colorless oil, IR (film) 1720, 1645, 910, 830 cm⁻¹; UV (EtOH) 206.5, 244.5 nm (ε 14500, 16400); CD, Δ ε₂₅₂ -0.54 (ε 2.5, EtOH); ¹H NMR δ 1.57, 1.62 (3H each, br.s), 1.68 (3H, s, 6-CH₃), 1.76—2.28 (4H, 4-H_{A.B}, 5-H_{A.B}), 5.12 (1H, m, 3-H), 5.17 (1H, dd, J=1.2, 10.0 Hz, 8-H_A), 5.23 (1H, dd, J=1.2, 17.5 Hz, 8-H_B), 6.07 (1H, dd, J=10.0, 17.5 Hz, 7-H), 7.52 (2H, aromatic protons) and 7.85 (2H, aromatic protons); MS m/z (%) 185, 183 [100, 98 (p-BrC₆H₄)+], 136 (43).

Isolation of Striatol (2). Fraction 3 (F-3) was rechromatographed on 10% AgNO₃–SiO₂ eluted with hexane–EtOAc (10:1) to yield colorless viscous oil (2, 70 mg), $[a]_{2}^{12}$ +49.5° (ε 1.2, CHCl₃); IR (film) 3400, 1640, 990, 800 cm⁻¹; ¹H NMR δ 0.87 (3H, d, J=6.5 Hz, 6-CH₃), 0.88 (3H, s, 5-CH₃), 1.29 (3H, s, 9-CH₃), 1.59 (3H, br.s, 4-CH₃), 5.09 (1H, dd, J=1.6, 10.5 Hz, 11-H_A), 5.26 (1H, dd, J=1.6, 17.5 Hz, 11-H_B), 5.41 (1H, m, 3-H) and 5.92 (1H, dd, J=10.5, 17.5 Hz, 10-H); ¹³C NMR δ 15.7 (q), 19.0 (q), 21.0 (q), 27.6 (q), 25.4 (t), 27.0 (t), 30.1 (t), 36.5 (t), 33.2 (d), 40.0 (s), 73.1 (s), 111.4 (t), 124.0 (d), 145.1 (d), 139.3 (s); MS m/z (%) 204 [17, (M

-18)+], 123 [100, (M-81)+], 81 (44). Found: m/z 222.1994, Calcd for $C_{15}H_{26}O$: M, 222.2004.

Benzoylation of Striatol (2). A solution of striatol 2 (10 mg) and p-bromobenzoyl chloride (10 mg) in dry pyridine (1 ml) was kept at room temperature for 2 d. The reaction mixture was treated in the usual way to give a residue, which was purified by SiO₂ column chromatography using hexane-EtOAc (15:1) to afford benzoate 2b (7 mg), colorless viscous oil, IR (film) 1720, 1645, 980, 800 cm⁻¹; UV (EtOH) 206, 244.5 nm (ε 16000, 18000); CD, $\Delta \varepsilon_{252} - 0.4$ (ε 2.7, EtOH), ¹H NMR δ 0.81 (3H, d, J=7.0 Hz, 6-CH₃), 0.81 (3H, s, 5-CH₃), 1.59 (3H, br.s, 4-CH₃), 1.69 (3H, s, 9-CH₂), 5.17 (1H, dd, I =1.6, 10.5 Hz, 11- H_A), 5.22 (1H, dd, J=1.6, 17.5 Hz, 11- H_B), 5.40 (1H, m, 3-H), 6.03 (1H, dd, J=10.5, 17.5 Hz, 10-H), 7.53 (2H, aromatic protons) and 7.85 (2H, aromatic protons); MS m/z (%) 185, 183 [100, 98 (p-BrC₆H₄CO)+], 204 (20), 123 (95), 81 (30).

Isolation of β-Monocyclonerolidol (3). Fraction 3' (F-3') was purified by column chromatography on 10% AgNO₃–SiO₂ using hexane–EtOAc (10:1) to give β-monocyclonerolidol (3, 70 mg), colorless viscous oil, $[a]_D^{25}$ +3.2° (c 0.66, CHCl₃); IR (film) 3400, 1645, 990, 890 cm⁻¹; ¹H NMR δ 0.84, 0.92 (3H each, s, gem-dimethyl at C-4), 1.27 (3H, s, 9-CH₃), 4.53 (1H, br.s, 14-H_A), 4.75 (1H, br.s, 14-H_B), 5.02 (1H, dd, J=1.6, 10.5 Hz, 11-H_A), 5.18 (1H, dd, J=1.6, 17.5 Hz, 11-H_B), and 5.91 (1H, dd, J=10.5, 17.5 Hz, 10H); ¹³C NMR δ 20.4 (q), 26.4 (q), 27.7 (q), 23.7 (t), 28.5 (t), 32.4 (t), 36.2 (t), 41.1 (t), 54.5 (d), 35.0 (s), 73.3 (s), 109.0 (t), 111.4 (t), 145.4 (d), 149.3 (s); MS m/z (%) 204 [25, (M-H₂O)+], 189 (40), 123 [100, (M-18-81)+], 109 (43), 93 (43), 81 (75). Found: m/z 222.1992. Calcd for C₁₅H₂₆O: M, 222.2004.

Hydrogenation of a-Ionone (13). The mixture of a-ionone 13 (500 mg) and Pd/C (50 mg) in EtOH containing 1.5% KOH was stirred under hydrogen atmosphere at room temperature for 4 h. The reaction mixture was filtered and the filtrate was neutralized with 5% aqueous HCl, diluted with H_2O , and extracted with ether. The ether layer was treated in the usual way to give a residue, which was chromatographed on SiO₂. Elution with CHCl₃ yielded dihydroionone 14 (190 mg), colorless oil, IR (film) 1710, 810 cm⁻¹; ¹H NMR δ 0.88, 0.93 (3H each, s, gem-dimethyl), 1.66 (3H, br.s__,CH₃), 2.12 (3H, s, COCH₃), 5.31 (1H, m, =/H); MS m/z (%) 194 (4, M+, $C_{13}H_{22}O$), 176 [23, (M-CH₃)+], 161 [17, (M-30)+], 136 [80, (M-15-43)+], 121 (100), 43 (84).

Acetalization of Dihydroionone 14. A solution of dihydroionone 14 (500 mg) in benzene (40 ml) containing ethylene glycol (6 ml) and TsOH (0.13 g) was stirred at reflux with a Dean-Stark trap for 12 h. The reaction mixture was washed with aqueous NaHCO₃ and H₂O, and dried over Na₂SO₄. The solvent was removed in vacuo to give a residue (580 mg), which was chromatographed on SiO₂. Elution with CHCl₃ gave acetal 15 (400 mg), colorless oil, IR (film) 1650, 1060, 860 cm⁻¹; ¹H NMR δ 0.86, 0.92 (3H each, s, gem-dimethyl) 1.30 (3H, s, O-CH₃), 1.66 (3H, br. s, CH₃), 3.90 (4H, s), 5.27 (1H, br.s, H); MS m/z (%) 238 (1, M+,C₁₅H₂₆O₂), 223 [2, (M-CH₃)+], 136 (25), 87 [100, (CH₃, O-)+], 43 (19).

Irradiation of Acetal 15. A solution of acetal 15 (2 g) in ether (270 ml) containing phenol (30 g) was irradiated with 450 W-Hg lamp for 3 h. The reaction mixture was passed an Al_2O_3 column by elution with ether. The ether fraction was evaporated under vacuum to give a residue (1.5 g), which was chromatographed on 10% $AgNO_3$ -SiO₂ to yield β -isomer 16 (550 mg) and starting material 15 (600 mg). β -Isomer 16,

colorless oil, IR (film) 1640, 1070, 890 cm⁻¹; ¹H NMR δ = 0.84, 0.94 (3H each, s, gem-dimethyl), 1.31 (3H, s, \square_{O}^{O}) CH₃), 3.92 (4H, s), 4.52, 4.73 (1H each, br.s, \Longrightarrow (\square_{H}^{H}); MS m/z (%) 238 (58, M⁺, C₁₅H₂₆O₂), 223 [100, (M-CH₃)⁺], 178 (73), 176 (93), 161 (97), 136 (98), 87 [99, (CH₃ \searrow_{O}^{O})⁺]. 43 (70).

Hydrolysis of Acetal 16. The mixture of acetal 16 (300 mg) and TsOH (20 mg) in THF (10 ml) and H₂O (1 drop) was kept at room temperature for 2 d. The reaction mixture was diluted with H₂O and extracted with CH₂Cl₂. The CH₂Cl₂ layer was treated in the usual way to afford a residue (250 mg), which was purified by SiO₂ column chromatography using CH₂Cl₂ to yield ketone 16a (190 mg), colorless oil, IR (film) 1710, 1640, 885 cm⁻¹; ¹H NMR δ 0.94, 0.96 (3H each, s, gem-dimethyl), 2.00 (3H, s, -COCH₃), 4.49, 4.74 (1H each, br. s, \rightleftharpoons H); MS m/z (%) 194 (2, M+, C₁₃H₂₂O), 176 [25, (M - CH₃)+], 161 (27), 136 (59), 121 (68), 43 (100).

Grignard Reaction of Ketone 16a. After a solution of ketone 16a (250 mg) in dry THF (5 ml) was added dropwise to a stirred solution of vinylmagnesium bromide (prepared from 3 g of vinyl bromide and 720 mg of Mg in 10 ml of dry THF) at room temperature, the mixture was heated under reflux for 3 h. The reaction mixture was then decomposed by addition of a saturated solution of NH₄Cl. The water layer was extracted with ether and the ether extract dried over Na₂SO₄. The solvent was evaporated under vacuum to give a residue, which was chromatographed on SiO₂. Elution with CHCl₃ yielded alcohol 17 (123 mg).

The IR, NMR, and mass spectra were found to be identical with those of β -monocyclonerolidol (3).

Isolation of Ptychanolide (4). The crude ptychanolide (F-2 and F-2') was recrystallized with hexane-EtOAc to yield colorless needles (4, 270 mg, 120 mg), mp 143-144 °C, $[a]_{D}^{24} + 23.2^{\circ} (c \ 0.47, \text{CHCl}_{3}); \text{IR (KBr) } 1780 \text{ cm}^{-1}; {}^{1}\text{H NMR}$ δ 0.79, 0.93 (3H each, s, 4-CH₃, 8-CH₃), 0.84 (3H, d, J=7.7Hz, 1-CH₃), 0.98 (3H, d, J=7.7 Hz, 5-CH₃), 1.3—1.4 (2H, $m, 2-H_A, 3-H_A$), 1.39 (1H, d, $J=13.6 \text{ Hz}, 7-H_A$), 1.71 (1H, m, $2-H_B$), 1.87 (1H, m, $3-H_B$), 1.99 (1H, d, J=13.6 Hz, $7-H_B$), 2.38 (1H, m, 1-H), 2.61 (1H, q, J=7.7 Hz), 3.55 (1H, d, J=2.4 Hz, 9-H and 5.52 (1H, d, J=2.4 Hz, 10-H); 13 C NMR δ 10.7 (q), 14.5 (q), 16.7 (q), 18.4 (q), 31.0 (t), 35.5 (t), 44.6 (t), 42.0 (d), 50.0 (d), 54.4 (s), 55.3 (s), 56.3 (s), 57.5 (d), 76.1 (d), 179.8 (s); MS m/z (%) 250 (6, M+), 235 [13, (M-CH₃)+], 205 (24), 109 (100). Found: m/z 250.1565. Calcd for $C_{15}H_{22}O_3$: M, 250.1564.

Hydrolysis of Ptychanolide (4). After 10% aqueous H₂SO₄ was added to a solution of ptychanolide 4 (20 mg) in acetone (2 ml) at 0 °C, the mixture was kept at room temperature for 3 h. The reaction mixture was diluted with H2O and extracted with CHCl₃. The CHCl₃ layer was washed with a saturated solution of NaHCO₃ and H₂O, and dried over Na₂SO₄. The solvent was removed under reduced pressure to give residue (24 mg), which was chromatographed on SiO₂. Elution with CHCl₃-MeOH (15:1) gave diol lactone 18 (17 mg), colorless powder, IR (CHCl₃), 3500, 1770 cm⁻¹; ¹H NMR δ 0.84 (3H, d, J=7.5 Hz, 1-CH₃), 0.80, 0.91 (3H each, s, 4-CH₃, 8-CH₃), 1.07 (3H, d, J=7.5 Hz, 5-CH₃), 2.67 (1H, q, J=7.5 Hz, 5-H), 4.13 (1H, d, J=2.0 Hz, 9-H) and 5.62 (1H, d, J=2.0 Hz, 10-H); MS m/z (%) 268 (trace, M⁺, $C_{15}H_{24}O_4$), 250 [4, (M- H_2O)+], 193 (100), 165 (98), 109 (97),

Reduction of Ptychanolide (4) with Lithium Aluminium Hydride. A solution of ptychanolide 4 (75 mg) in dry ether (2 ml) was added to an ice-cooled solution of LAH (100 mg) in dry ether

(4 ml). After addition, the reaction mixture was stirred at room temperature for 2 h followed by the usual work-up afforded a residue. The residue was chromatographed on SiO₂ using CHCl₃-MeOH (15:1) to give triol 19 (75 mg). recrystallization of triol 19 from MeOH yielded colorless plates, mp 118—119 °C, $[\alpha]_D^{28}$ -40.0° (c 1.04, CHCl₃); IR (CHCl₃) 3420, 3250 cm⁻¹; ¹H NMR δ 0.69, 0.82 (3H each, s, 4-CH₃, 8-CH₃), 0.81 (3H, d, J=7.7 Hz, 1-CH₃), 1.16 (3H, d, J=7.7 Hz, 5-CH₃), 1.10 (1H, d, J=13.6 Hz, 7-H_A), 1.42 (1H, d, J=13.6 Hz, 7-H_B), 2.15 (1H, q, J=7.7 Hz, 5-H), 2.65, 3.20 (1H each, m, OH), 3.26 (1H, br.s, OH), 3.64, 3.84 (1H each, AB type, J = 14.4 Hz, $-C\underline{H}_2OH$), 3.72 (1H, dd, J =7.2, 14.4 Hz, -CH-HCH-OH), 3.76 (1H, dd, J=4.0, 14.4 Hz, -CH-HC<u>H</u>-OH) and 3.92 (1H, dd, J=4.0, 7.2 Hz, -C<u>H</u>- $(OH)-CH_2OH)$; MS m/z (%) 225 [75, $(M-31)^+$], 207 (80), 189 (58), 109 (100). Found: C, 69.98; H, 11.04%. Calcd for C₁₅H₂₈O₃: C, 70.27; H, 11.01%.

Oxidation of Triol 19 with NaIO₄. A solution of NaIO₄ (25 mg) in H₂O (1 ml) was added to a solution of triol 19 (20 mg) in MeOH (1 ml) and the mixture was stirred at room temperature for 3 h. The reaction mixture was diluted with H₂O and extracted with ether. The residue after usual treatment was chromatographed on SiO₂ and elution with CHCl₃-MeOH (25 : 1) afforded aldehyde 20 (16 mg), colorless oil, $[a]_{\rm D}^{25}$ -3.3° (c 0.42, CHCl₃); IR (film) 3560, 3450, 1715 cm⁻¹; ¹H NMR δ 0.81 (6H, s, 4-CH₃, 8-CH₃), 0.89 (3H, d, J=7.5 Hz, 1-CH₃), 0.92 (3H, d, J=7.5 Hz, 5-CH₃), 3.55, 3.94 (1H each, d, J=11.0 Hz, CH₂OH) and 9.69 (1H, s, CHO); MS m/z (%) 224 (2, M+), 206 [15, (M-H₂O)+], 195 (51), 181 (100), 109 (90). Found: m/z 224.1771. Calcd for C₁₄H₂₄O₂: M, 224.1768.

Oxidation of Aldehyde (20) with MCPBA. A solution of aldehyde 20 (16 mg) in CH₂Cl₂ (1 ml) was added to a solution of MCPBA (20 mg) and the mixture stirred at room temperature for a day. Then the reaction mixture was treated with Al₂O₃ for 30 min. Al₂O₃ was removed by filtration and the solvent was evaporated to give a residue, which was purified by column chromatography on SiO₂. Elution with CHCl₃-MeOH (15:1) gave diol 21 (12 mg), colorless oil, $[a]_{23}^{23}-18.4^{\circ}$ (c 0.57, CHCl₃); IR (film) 3325, 1050 cm⁻¹; ¹H NMR δ 0.88 (3H, d, J=7.5 Hz, 1-CH₃), 0.89, 0.93 (3H each, s, 4-CH₃, 8-CH₃), 0.90 (3H, d, J=7.5 Hz, 5-CH₃), 1.47 (1H, d, J=13.5 Hz, 7-H_A), 1.92 (1H, d, J=13.5 Hz, 7-H_B), 3.42 (2H, AB q, J=11.0 Hz, -CH₂OH); MS m/z (%) 212 (3, M+, C₁₃H₂₄O₂), 194 [5, (M-H₂O)+], 181 [100, (M-31)+], 163 (62), 137 (53), 109 (97).

Oxidation of Diol 21 with NaIO₄. A solution of NaIO₄ (20 mg) in H₂O (1 ml) was added to a MeOH (1.5 ml) solution of diol 21 (17 mg) and the mixture was stirred at room temperature for a day. The reaction mixture was diluted with H₂O and extracted with hexane. The solvent was removed under vacuum to give a residue (15 mg), which was chromatographed on SiO₂. Elution with CH₂Cl₂ yielded ketone 22 (9 mg), colorless oil, CD, $\Delta \varepsilon_{295} + 2.6$ (c 0.0115, MeOH); IR (film) 1740 cm⁻¹; ¹H NMR δ 0.81, 0.87 (3H each, s, 4-CH₃, 8-CH₃), 0.93, 0.95 (3H each, d, J=7.5 Hz, 1-CH₃, 5-CH₃), 1.55 (1H, d, J=13.5 Hz, 7-H_A), 2.32 (1H, d, J=13.5 Hz, 7-H_B) and 2.27 (1H, q, J=7.5 Hz, 5-H); MS m/z (%) 180 (82, M⁺, C₁₂H₂₀O), 165 [33, (M-CH₃)⁺], 123 (46), 109 (100), 82 (88).

Reduction of Ketone 22 with Lithium Aluminium Hydride. A mixture of ketone 22 (9 mg) and LAH (20 mg) in dry ether (3 ml) was stirred at room temperature for 2 h. The reaction mixture was treated in the usual way to give a residue (9 mg), which was purified by column chromatography on SiO₂. Elution with CHCl₃ gave a minor alcohol 24 (2 mg), colorless oil, IR (film) 3430 cm⁻¹; ¹H NMR δ 0.84, 0.90 (3H each, s, 4-CH₃, 8-CH₃), 0.84 (3H, d, J=7.7 Hz, 1-CH₃), 0.91 (3H, d,

J=7.7 Hz, 5-CH₃), 1.42 (1H, dd, J=5.4, 13.6 Hz, 7-H_A), 1.95 (1H, dd, J=7.2, 13.6 Hz, 7-H_B), 1.77 (1H, m, 1-H), 1.87 (1H, dq, J=7.2, 7.7 Hz, 5-H) and 4.20 (1H, dt, J=5.4, 7.2 Hz, 6-H). Successive elution with the same solvent gave a major alcohol 23 (5 mg), colorless oil, IR (film) 3450 cm⁻¹; ¹H NMR δ 0.75 (6H, s, 4-CH₃, 8-CH₃), 0.84 (3H, d, J=7.7 Hz, 1-CH₃), 0.91 (3H, d, J=7.7 Hz, 5-CH₃), 1.49 (1H, dd, J=7.7, 13.6 Hz, 7-H_A), 1.74 (1H, dq, J=9.4, 7.7 Hz, 5-H), 1.79 (1H, dd, J=7.7, 13.6 Hz, 7-H_B), 2.01 (1H, tq, J=7.2, 8.1 Hz, 1-H), 3.77 (1H, dt, J=9.4, 7.7 Hz, 6-H); MS m/z (%) 164 [32, (M−H₂O)+], 149 [100, (M−18−15)+].

Conversion of Ketone 22 into Hydrocarbon 25. ketone 22 (10 mg) in 1,2-ethanedithiol (0.1 ml) and BF₃-Et₂O (2 drops) was kept at room temperature for 2 h. The reaction mixture was poured into ice-water, and extracted with ether. The ether layer was washed with a saturated solution of NaHCO3 and H2O, and dried over Na2SO4. The solvent was removed under vacuum to give a residue, which was desulfurized with Raney Ni (W-4, 50 mg) in EtOH (1.5 ml) under reflux for 2 h. The reaction mixture was filtered and the filtrate was diluted with H₂O, and extracted with pentane. The solvent was removed with N2 gas to give a colorless residue, which was subjected to GC separation to yield hydrocarbon 25 (4 mg), colorless oil, ¹H NMR δ 0.73 (6H, s, 4-CH₃, 8-CH₃), 0.83 (6H, d, J=7.7 Hz, 1-CH₃, 5-CH₃), 1.27 (2H, m, 3-H_A) $7-H_A$), 1.30 (2H, m, $2-H_A$, $6-H_A$), 1.50 (2H, m, $3-H_B$, $7-H_B$), 1.73 (2H, m, 2-H_B, 6-H_B), and 1.85 (2H, m, 1-H, 5-H); 13 C NMR δ 15.0, 18.0, 31.7, 37.5, 44.4, 53.7; MS m/z (%) 166 $(9, M^+, C_{12}H_{22}), 151 [51, (M-CH_3)^+], 109 (61), 84 (100).$

We wish to thank Dr. Jiro Hasegawa of Kyoto Univ. for identification of *Ptychanthus striatus* (Lehm. et Lindenb.) Nees, to Professor Yoshinori Asakawa of Tokushima Bunri Univ. for supply of deoxopinguisone and Professor Koji Nakanishi of our institute director for many discussions.

References

1) V. Benesova, Z. Samek, V. Herout, and F. Sorm, Coll. Czech. Chem. Comm., 34, 582 (1969); H. Knoche, G. Ourisson, G. W. Perold, J. Foussereau, and J. Maleville, Science, 166, 239 (1969); S. Huneck, J. Hattori Bot. Lab., 43, 468(1972); N. H. Andersen, B. Shunk, and C. R. Costin, Experimentia, 29, 645 (1973); S. Hayashi and A. Matsuo, Hikobia, 7, 125 (1975); C. Suire, Rev. Bryol. Lichen., 41, 105 (1975); Y. Asakawa, T. Takemoto, M. Toyota, and T. Aratani, Tetrahedron Lett., 1977, 1407; "Progress in Phytochemistry," ed by L. Reinhold, J. B. Harborne, and T. Swain, Pergamon Press, London (1978),

- Vol. 5, p. 181.
- 2) S. M. Krutov, Z. Samek, V. Benesova, and V. Herout, *Phytochemistry*, 12, 1405 (1973).
- 3) Y. Asakawa, C. Suire, M. Toyota, N. Tokunaga, T. Takemoto, S. Hattori, and M. Mizutani, J. Hattori Bot. Lab., 46, 77 (1980).
- 4) R. Takeda, R. Mori, and Y. Hirose, Chemistry Lett., 1982, 1625.
- 5) R. Takeda, H. Naoki, T. Iwashita, and Y. Hirose, Tetrahedron Lett., 22, 5307 (1981).
- 6) The present result was confirmed by NOE difference spectrum.
- 7) C. L. Kissel and B. Rickborn, *J. Org. Chem.*, **37**, 2060 (1972).
- 8) N. Harada, J. Iwabuchi, Y. Yokota, H. Ueda, and K. Nakanishi, J. Am. Chem. Soc., 103, 5590 (1981).
- 9) N. C. Gonnella, K. Nakanishi, V. S. Martin, and K. B. Sharpless, J. Am. Chem. Soc., 104, 3775 (1982).
- 10) P. J. Kropp and H. J. Krauss, J. Am. Chem. Soc., 89, 5199 (1967); S. G. Levine and B. Gopalakrishnan, Tetrahedron Lett., 1979, 699.
- 11) Y. Asakawa, N. Tokunaga, M. Toyota, T. Takemoto, and C. Suire, J. Hattori Bot. Lab., 45, 395 (1979).
- 12) B. M. Howard and W. Fenical, Tetrahedron Lett., 1976,
- 13) G. Cimino, S. D. Stefano, A. Gueriero, and L. Minale, Tetrahedron Lett., 1975, 3723.
- 14) G. Cimino, S. D. Stefano, A. Gueriero, and L. Minale, Tetrahedron Lett., 1975, 1417, 1425; R. J. Capon, E. L. Ghisalberti, and P. R. Jefferies, Aust. J. Chem., 34, 1775 (1981).
- 15) P. Main, S. E. Hull, L. Lessinger, G. Germain, J. P. Declercq, and M. M. Woolfson (1978). MULTAN78, A System of Computer Pograms for the Automatic Solution of Crystal Structures from X-ray Diffraction Data. University of York, England, and Louvain, Belgium.
- 16) "International Tables for X-Ray Crystallography," Kynoch Press, Birmingham (1974), Vol. IV.
- 17) The Universal Crystallographic Computing System (1979). Library of Programs, Osaka University, Computing Center.
- 18) W. Moffitt, R. B. Woodward, A. Moscowitz, W. Klyne, and C. Djerassi, J. Am. Chem. Soc., 83, 4013 (1961); "Natural Products Chemistry," ed by K. Nakanishi, T. Goto, S. Ito, S. Natori, and S. Nozoe, Kodansha, Tokyo and Academic Press, N. Y., London (1974), Vol. 1, p. 28.
- 19) A. Corbella, P. Gariboldi, G. Jommi, F. Orsini, A. DeMarco, and A. Immirzi, J. Chem. Soc., Perkin Trans. 1, 1974, 1875.